www.nature.com/mp

# **PERSPECTIVE** Myelination of parvalbumin interneurons: a parsimonious locus of pathophysiological convergence in schizophrenia

J Stedehouder and SA Kushner

Schizophrenia is a debilitating psychiatric disorder characterized by positive, negative and cognitive symptoms. Despite more than a century of research, the neurobiological mechanism underlying schizophrenia remains elusive. White matter abnormalities and interneuron dysfunction are the most widely replicated cellular neuropathological alterations in patients with schizophrenia. However, a unifying model incorporating these findings has not yet been established. Here, we propose that myelination of fast-spiking parvalbumin (PV) interneurons could be an important locus of pathophysiological convergence in schizophrenia. Myelination of interneurons has been demonstrated across a wide diversity of brain regions and appears highly specific for the PV interneuron subclass. Given the critical influence of fast-spiking PV interneurons for mediating oscillations in the gamma frequency range (~30–120 Hz), PV myelination is well positioned to optimize action potential fidelity and metabolic homeostasis. We discuss this hypothesis with consideration of data from human postmortem studies, *in vivo* brain imaging and electrophysiology, and molecular genetics, as well as fundamental and translational studies in rodent models. Together, the parvalbumin interneuron myelination hypothesis provides a falsifiable model for guiding future studies of schizophrenia pathophysiology.

Molecular Psychiatry (2017) 22, 4-12; doi:10.1038/mp.2016.147; published online 20 September 2016

Schizophrenia is a chronically debilitating psychiatric disorder with a lifetime prevalence of ~1%.<sup>1</sup> Patients with schizophrenia classically exhibit a constellation of positive, negative and cognitive symptoms.<sup>2</sup> Although many theories have been proposed, the precise neurobiological mechanism underlying schizophrenia has remained elusive. The most widely described models have been the dopamine<sup>3</sup> and glutamate hypotheses,<sup>4</sup> although in recent years models regarding interneuron dysfunction<sup>5</sup> and myelination abnormalities<sup>6</sup> have gained increasing support.

In this Perspective, we hypothesize that previous observations of interneuron dysfunction and myelination abnormalities in schizophrenia might converge on the altered myelination of fast-spiking parvalbumin (PV) interneurons. First, we summarize the major evidence supporting interneuron dysfunction and myelination abnormalities in schizophrenia. Next, we summarize electron microscopy and immunofluorescence studies that convincingly demonstrate interneuron myelination, which frequently occurs on fast-spiking PV interneurons. Finally, we discuss how impairments in myelination of PV interneurons could lead to consequent abnormalities in gamma synchronization and ultimately give rise to the symptoms which define schizophrenia.

# PARVALBUMIN INTERNEURON DYSFUNCTION IN SCHIZOPHRENIA

Deficits in GABAergic signaling have been widely proposed as a fundamental pathophysiological mechanism underlying schizophrenia.<sup>7</sup> More specifically, several recent lines of evidence including human postmortem studies, genetics and *in vivo* electrophysiological recordings in patients and translational mouse models have identified fast-spiking PV interneurons as the major GABAergic cell-type affected in schizophrenia (Table 1).

Expression of GAD67—the predominant gamma-aminobutyric acid (GABA) synthesizing enzyme—has consistently been found to be reduced at both the messenger RNA and protein levels in several brain regions of patients with schizophrenia, a finding that has been well controlled for confounding factors.<sup>8–14</sup> Downregulation of GAD67 messenger RNA has been reported in ~30% of dorsolateral prefrontal cortex interneurons<sup>15,16</sup> and entirely undetectable in ~50% of PV interneurons.<sup>17</sup> Expression of PV messenger  $RNA^{18-20}$  and protein<sup>21</sup> is also reduced in schizophrenia, while the neuronal density of cortical PV interneurons is unchanged<sup>22-25</sup> (but see also ref. 52). Since the expression of both PV and GAD67 are experience-dependent<sup>26</sup>—and GAD67 and PV expression are highly correlated<sup>26</sup>—their shared downregulation suggests a functional impairment of fast-spiking interneurons.<sup>2</sup> Morphologically, PV cell inputs onto pyramidal neurons have no discernible alterations,<sup>21</sup> suggesting a primary functional abnormality of PV interneurons. Consistent with these neuropathological findings, in vivo positron emission tomography (PET) imaging has demonstrated widespread alterations of cortical GABA transmission in schizophrenia, a finding that was most prominent in the subset of patients who were antipsychotic-naïve.<sup>28</sup> Altogether, these results provide compelling evidence of cortical PV interneuron dysfunction in schizophrenia.

PV interneurons are essential in generating cortical oscillations in the gamma range (~30–120 Hz), mediated by synchronized inhibition of large pyramidal cell ensembles.<sup>29,30</sup> Through rhythmic perisomatic inhibition onto surrounding pyramidal cells, synchronous ensembles of PV cells evoke high-frequency gamma oscillations in the cerebral cortex.<sup>31–33</sup> Gamma synchrony has been shown to function critically across a range of cognitive

Department of Psychiatry, Erasmus University Medical Center, Rotterdam, The Netherlands. Correspondence: Dr SA Kushner, Department of Psychiatry, Erasmus University Medical Center, Wytemaweg 80, 3015 CN Rotterdam, The Netherlands.

E-mail: s.kushner@erasmusmc.nl

Received 20 December 2015; revised 9 July 2016; accepted 13 July 2016; published online 20 September 2016

	Interneuron dysfunction	Myelination abnormalities
Schizophrenia age of onset	Maturation of PV cells <sup>2</sup>	Peak of myelination <sup>63</sup>
. 5	Emergence of high-frequency oscillations <sup>35</sup>	Development of frontal gray matter oligodendrocytes <sup>135</sup>
Postmortem findings	PV mRNA and protein decreased <sup>18-21</sup>	Abnormal myelin/oligodendrocyte gene expression64-71
5	GAD67 mRNA and protein decreased <sup>8-14</sup>	Lower oligodendrocyte numbers <sup>73–83</sup>
	Transcriptional changes in PV cells <sup>150</sup>	Ultrastructural abnormalities <sup>89,90</sup>
	······································	Transcriptional changes in oligodendrocyte lineage cells <sup>83</sup>
Human <i>in vivo</i> findings	Activity-dependent EEG abnormalities <sup>5,34,35</sup> a	Lower FA values on DTI <sup>44–58</sup> a
i annan <i>in</i> tire innanige	MRS-based GABA impairments <sup>28</sup>	
Genetic support	CNVs <sup>36</sup>	GWAS common variants in myelin/oligodendrocyte gene sets <sup>92-9</sup>
denetic support		GWAS common variants enriched in mature oligodendrocytes <sup>96</sup>

functions, including working memory and attention,<sup>34</sup> with wellreplicated abnormalities in schizophrenia.<sup>5,35</sup> Abnormalities in other frequency bands such as theta and alpha have also been reported in schizophrenia, but the neural mechanisms underlying these frequencies remain less well understood.<sup>35</sup>

episode, drug-naïve patients.

Electroencephalographic studies in schizophrenia have shown a reduced amplitude and impaired phase locking of gamma band activity over frontal areas while assessing working memory and executive functioning tasks.<sup>35</sup> Although some studies have observed concurrent increases in gamma band activity at rest, this finding has been less well replicated.<sup>35</sup> Taken together, impairments of *in vivo* gamma oscillations in patients with schizophrenia are highly consistent with the PV interneuron abnormalities observed by postmortem histopathology.

The classical onset of schizophrenia occurs within a relatively narrow window of neurodevelopment, between ~ 18 and 25 years of age.<sup>2</sup> This late adolescent age of onset has often been attributed to the ongoing functional maturation of the brain during this neurodevelopmental critical period.<sup>2</sup> Specifically in late adolescence, rates of synaptic pruning and myelination become asymptotic for which impairments in these processes have been linked to the disease onset.<sup>2</sup> Notably, maturation of gamma band synchrony also occurs during late adolescence<sup>35</sup> which coincides developmentally with the clinical onset of schizophrenia.<sup>1</sup>

In addition to *in vivo* brain imaging, electroencephalographic recordings and postmortem histopathology, molecular genetic studies of schizophrenia have also revealed an important contribution of interneuron dysfunction to the pathophysiology of schizophrenia. A recent genetic study of copy number variation has now provided causal evidence for GABAergic dysfunction in the etiology of schizophrenia.<sup>36</sup> In this study, Pocklington et al. performed a functional gene set analysis for enriched biological mechanisms using a large schizophrenia case-control dataset and found that copy number variations were significantly enriched in cases for genes responsible for inhibitory neurotransmission (in particular the GABA<sub>A</sub> receptor complex), glutamatergic neurotransmission, long-term synaptic plasticity and associative learning. The genetic variant with the highest known risk for schizophrenia is the 22q11 microdeletion which has a penetrance of ~40%.  $^{\rm 37,38}$  Transgenic mouse models have been generated to investigate the underlying neurobiology conferred by 22q11 microdeletion. Df(16)A mice harboring a 27-gene microdeletion syntenic to a 1.5 Mb region of human 22g11.2 exhibit similar brain abnormalities as found in human 22q11 microdeletion carriers, including cortico-cerebellar, cortico-striatal and cortico-limbic circuits.<sup>39</sup> Moreover, multiple different mouse models of 22q11 microdeletion have replicated a cell-type specific impairment in PV interneurons and disrupted local synchrony of neural activity, consistent with the deficit in gamma oscillations observed in schizophrenia.<sup>40-4</sup>

Evidence for interneuron dysfunction in schizophrenia has also been supported by a wide variety of non-genetic rodent models.<sup>43</sup> The major examples include pharmacological NMDA receptor antagonism and neurodevelopmental immunological challenge, both of which consistently exhibit synaptic and network abnormalities reminiscent of schizophrenia pathophysiology. Specifically, these studies have identified electrophysiological changes in local microcircuit connectivity and synaptic plasticity, with alterations in excitation/inhibition balance and gamma band synchronization.

Taken together, the combination of genetic, postmortem, and *in vivo* electrophysiological and functional imaging results from human clinical studies of schizophrenia converge with translational rodent modeling to identify fast-spiking PV interneuron dysfunction as a major pathophysiological mechanism underlying schizophrenia etiology.

# **MYELINATION ABNORMALITIES IN SCHIZOPHRENIA**

Independent of PV interneuron alterations, myelination abnormalities have also been extensively implicated in schizophrenia through both in vivo brain imaging and postmortem assessments (Table 1). Numerous diffusion tensor imaging studies have been published for schizophrenia (reviewed in ref. 6), of which the overwhelming consensus has been the association of schizophrenia with globally decreased fractional anisotropy. Notably, the decrease in fractional anisotropy appears to become more severe with increasing age and illness duration.<sup>44</sup> Many of the early brain imaging studies of schizophrenia were performed in cohorts with extensive histories of psychotropic medication, inpatient hospitalization, smoking and medical co-morbidities, which could have a confounding deleterious influence on white matter integrity. Thus, an important question has been whether myelination abnormalities are already present in drug-naïve patients with first-episode schizophrenia who have never received psychotropic medication. Recently, several diffusion tensor imaging studies have been performed in such cohorts, 44-58 holding the potential to directly evaluate these potential confounders. Indeed, across a range of different methodologies, studies of drug-naïve firstepisode schizophrenia have consistently demonstrated similar, albeit less severe, myelination abnormalities as observed in chronic illness. Importantly, these studies confirm that a global impairment of myelin integrity is already present at the time of the initial clinical onset of psychotic symptoms in schizophrenia. Accordingly, these findings support a model by which myelination abnormalities function critically in the pathophysiology of schizophrenia.

The late adolescent age of onset for schizophrenia closely overlaps with the maturation of prefrontal cortex myelination.<sup>59</sup> The time course of myelination in humans has been elegantly detailed through longitudinal *in vivo* imaging and postmortem cross-sectional studies demonstrating rapid early postnatal white matter development in the first 12 months,<sup>60</sup> followed by a slower but steady increase until late adolescence.<sup>61,62</sup> Comparative mammalian evolutionary studies have demonstrated that humans exhibit a particularly extended neurodevelopmental time course of neocortical myelination.<sup>63</sup> Although myelination in humans peaks in late adolescence, for non-human primates and rodents the peak of myelination occurs significantly earlier in development.<sup>63</sup> Together, the current best evidence demonstrates that the onset of schizophrenia closely coincides with the peak of myelination in human brain development.

In addition to the well-replicated finding of *in vivo* white matter abnormalities in schizophrenia, postmortem gene expression analyses have also identified alterations in myelination regulatory pathways. Several studies have reported a broad reduction in the expression of genes with demonstrated function in the oligodendrocyte lineage.<sup>64–71</sup> Using microarray-based transcriptome analysis with quantitative PCR validation, abnormalities in oligodendrocyte lineage genes have been found in both frontal white and gray matter,<sup>64,65,68</sup> subcortical regions,<sup>66,69</sup> occipital cortex<sup>70</sup> and temporal cortex.<sup>71</sup> The alignment between *in vivo* brain imaging findings and postmortem gene expression analyses is highly consistent with a central importance of myelination abnormalities in schizophrenia pathophysiology. Notably, many of the same oligodendrocyte and myelination genes found to be altered in schizophrenia also exhibit consistent increases during normal brain development precisely during adolescence,<sup>72</sup> again consistent with the association between the late adolescent age of onset in schizophrenia and the peak of myelination.

Compared with the abundance of brain imaging and gene expression studies of myelination and oligodendrocytes, postmortem stereological analysis of oligodendrocyte lineage cell types are scarce. From the few studies that have been performed, stereological quantification of myelinating oligodendrocytes have revealed widespread reductions in schizophrenia (Table 1).73-83 Reductions in oligodendrocyte numbers have been shown in the white and gray matter of BA9,<sup>73–76,83</sup> white and gray matter of BA10,<sup>77,78</sup> posterior hippocampal subregion CA4,<sup>79</sup> internal capsule,<sup>80</sup> nucleus basalis<sup>81</sup> and anterior thalamic nucleus.<sup>82</sup> In contrast, oligodendrocyte numbers appear unchanged within the substantia nigra,<sup>84</sup> callosal genu<sup>85</sup> and subgenual cingulum.<sup>85</sup> Furthermore, one study failed to find differences in oligodendrocyte number within any subregion of the hippocampus.86 In addition, a few studies have reported seemingly paradoxical increases in the number of myelinating oligodendrocytes in frontal white matter<sup>87</sup> and basolateral amygdala.<sup>88</sup> Although caution is warranted given the limited number of studies and differences in methodology, the emerging picture is one of small but consistent reductions of myelinating oligodendrocytes in schizophrenia (~14% reduction<sup>73-82</sup>). However, an important unanswered question is whether the observed reduction of myelinating oligodendrocytes is cell-type specific or also extends to other less differentiated cell types within the oligodendrocyte lineage.

A recent study is the first to report a stereological analysis of oligodendrocyte precursors cells (OPCs),<sup>83</sup> also known as neuron-glial antigen 2 (NG2) cells due to their abundant expression of the NG2 protein. The number of frontal white matter OPCs were unchanged while the total population of oligodendrocyte lineage cells was reduced, thereby suggesting that the reduction in oligodendrocyte lineage cells occurs predominantly in more differentiated cell types. Furthermore, oligodendrocyte lineage-specific transcriptome analysis and immunohistochemical labeling independently suggested an impairment of OPC differentiation towards mature oligodendrocytes. Given that OPCs are the exclusive progenitor cell population of myelinating oligodendrocytes, more knowledge of the

Molecular Psychiatry (2017), 4-12

regulation and function of OPCs in schizophrenia would better clarify whether the observed reductions in myelinating oligodendrocytes are a consequence of abnormalities that have occurred upstream in the myelination lineage or the consequence of a downstream cell-type specific loss of myelinating oligodendrocytes.

Two studies have examined myelination at the ultrastructural level in schizophrenia. The major findings involved myelinated axons and oligodendrocytes, in frontal cortex white and gray matter.<sup>89,90</sup> The observed pathological features included alterations in the morphology of the myelin sheath and the frequency of axonal degeneration within morphologically intact myelin segments. Notably, the effect sizes were larger in gray matter compared with white matter regions.<sup>89,90</sup>

With regard to *in vivo* and postmortem findings, the possibility remains that the observed myelination abnormalities in schizophrenia could result from primary and/or secondary disturbances of neuronal signaling.<sup>91</sup> Therefore, genetic studies provide a unique opportunity to investigate etiological mechanisms of schizophrenia while avoiding the potential confounds of antipsychotic medication and secondary disease effects. Notably, recent studies have shown using genome-wide association study (GWAS) data that myelination/oligodendrocyte gene sets are significantly associated with both the risk of schizophrenia<sup>92–94</sup> and the severity of deficits in white matter integrity.<sup>95</sup> Moreover, the most recent GWAS results for schizophrenia exhibited a significant enrichment of genes expressed in mature oligodendrocytes,<sup>96</sup> together suggesting a convergence of common variant risk on myelination.

Although important questions remain unanswered, GWAS results implicating myelination as an etiological mechanism, *in vivo* imaging demonstrating well-replicated myelination abnormalities, human postmortem histopathology showing replicated decreases in the number and ultrastructure of oligodendrocytes, and gene profiling studies demonstrating replicated changes in oligodendrocyte expression, together provide compelling support for myelination as a major pathophysiological mechanism of schizophrenia.

# MYELINATION OF PARVALBUMIN INTERNEURONS

An increasing number of studies has revealed the unexpectedly extensive myelination of GABAergic interneurons (Table 2), predominantly fast-spiking PV basket cells (Table 3), in cortical gray matter and other regions throughout the brain.<sup>97–123</sup> Myelination of cortical GABAergic basket cells was first reported over 30 years ago in the cat visual cortex by electron microscopy.<sup>98–100</sup> In non-human primates, GABAergic axons in layers III–V are myelinated in sensorimotor<sup>106–108</sup> and temporal<sup>102</sup> cortices, a finding that had already been hinted at several years earlier.<sup>105</sup> Myelinated GABAergic interneurons were subsequently identified in the cat superior colliculus,<sup>118</sup> as well as in the red nucleus<sup>110</sup> and hypoglossal nucleus<sup>109</sup> of the monkey. Although the relative distribution of myelination across interneuron subtypes has not yet been quantitatively determined, a consistent qualitative observation has been that a high proportion of the GABA-labeled terminals of myelinated axons exhibit localized somatic targeting suggestive of basket cells.<sup>106</sup> Moreover, direct ultrastructural evidence for basket cell myelination has also been demonstrated in visual cortex of cat<sup>98,99</sup> and rat.<sup>112</sup>

Basket cells have been reported to be myelinated in occipital<sup>112</sup> and somatosensory<sup>114</sup> cortex of the rat. PV-immunoreactive myelinated neurons have been identified in the rat entorhinal cortex,<sup>119</sup> hippocampus<sup>122</sup> and striatum.<sup>123</sup> In the rat entorhinal cortex, myelinated PV axons were found extensively across all cortical layers, interspersed with unmyelinated axonal segments.<sup>119</sup> Furthermore, myelinated GABAergic neurons have been identified in the rodent hippocampus,<sup>111,117</sup> thalamus<sup>113,120</sup>

Study	Species	Brain region	Technique	Conclusion
Somogyi et al. <sup>100</sup>	Cat	Visual cortex	Electron microscopy	Presence of single myelinated GAD+ cells
Mize et al. <sup>118</sup>	Cat	Superior colliculus	Electron microscopy	Presence of myelinated GABAergic neurons
Ong et al. <sup>102</sup>	Human	Frontal cortex	Electron microscopy	Presence of several myelinated GAT-1 axons
Ong et al. <sup>102</sup>	Monkey	Temporal cortex	Electron microscopy	Presence of several myelinated GAT-1 axons
Hendry et al. <sup>106</sup>	Monkey	Sensory-motor cortex	Electron microscopy	Presence of several myelinated layers III–V GABAergic neurons
DeFelipe et al. <sup>107</sup>	Monkey	Somatosensory cortex	Electron microscopy; [ <sup>3</sup> H]GABA tracing	Presence of several myelinated GABAergic neurons
DeFelipe et al. <sup>108</sup>	Monkey	Sensorymotor cortex	Electron microscopy	Presence of several myelinated Layers III–V GABAergic neurons
Takasu <i>et al.</i> <sup>109</sup>	Monkey	Hypoglossal nucleus	Electron microscopy	Presence of several myelinated GABAergic neurons
Ralston et al. <sup>110</sup>	Monkey	Red nucleus	Electron microscopy	Presence of several myelinated GABAergic neurons
Jinno <i>et al</i> . <sup>111</sup>	Rat	Hippocampus	Single cell tracing; immunofluorescence	Presence of several myelinated GABAergic projection neurons
De Biasi et al. <sup>113</sup>	Rat	Thalamus	Electron microscopy	Presence of a few myelinated GABAergic axons
Conti <i>et al</i> . <sup>114</sup>	Rat	Cortex	Electron microscopy	Presence of several myelinated GAT-2 positive axons
Roberts et al. <sup>121</sup>	Rat	Inferior colliculus	Electron microscopy	Presence of several myelinated GABAergic neurons
Sawyer et al. <sup>120</sup>	Rat	Thalamus	Electron microscopy; light microscopy	Presence of several myelinated GABAergic neurons

and inferior colliculus,<sup>121</sup> although these studies were performed largely without interneuron subtype-specific labeling. However in one notable exception, myelinated rat hippocampal GABAergic neurons were confirmed as PV interneurons.<sup>117</sup> Moreover, the vast majority of septohippocampal PV, but not cholinergic, fibers are myelinated.<sup>115,116,124</sup>

Recently, a combinatorial study using array tomography and fluorescence microscopy found a large abundance of myelinated PV axons in adult mouse somatosensory cortex.<sup>125</sup> Here, the authors found that 25–50% of all cortical myelinated axons are GABAergic, and that nearly all of these are PV-expressing interneurons, a finding that has been independently observed in the adolescent mouse visual cortex.<sup>97</sup> Notably, although myelin thickness was similar between GABAergic and non-GABAergic axons, myelinated GABAergic axons had a higher average g-ratio (ratio between the inner axonal diameter and the outer diameter of the myelin sheath), shorter internode length, and shorter nodes of Ranvier than non-GABAergic axons.<sup>125</sup>

Few studies have reported attempts to examine myelination of interneurons in human cortex. Myelination of PV cells in the human hippocampus<sup>103</sup> and claustrum<sup>104</sup> has been confirmed by electron microscopy. Furthermore, myelinated GABAergic<sup>102</sup> interneurons have been incidentally observed in the human frontal cortex, including with PV subtype specification.<sup>101</sup> Thus, although sparsely documented, PV interneuron myelination appears to be widespread throughout the brain and evolutionarily conserved among mammals. More detailed and comprehensive studies are required to quantify the relative proportion of myelinated PV interneurons, their developmental time course of myelination compared to pyramidal neurons, subcellular distribution of segmental myelination, and brain region distribution, as well as the functional neurophysiological implications of interneuron myelination.

Notably, we have not found any report demonstrating myelination of cortical somatostatin or neuropeptide Y interneurons, despite numerous electron microscopic studies in a variety of mammalian species,<sup>126–129</sup> thereby suggesting a high specificity for the PV subclass of GABAergic interneurons. In contrast, non-PV interneuron myelination has been sporadically reported in subcortical regions, for example, in sparse small-diameter axons of the rat internal capsule<sup>130</sup> and in the cat claustrum.<sup>131</sup> This suggests that at least within the cerebral cortex, PV cells are the predominant myelinated interneuron subtype while in subcortical brain regions the cell-type distribution of myelinated interneurons may be less strict.

Recently, it has been shown that PV interneurons establish direct functional soma-targeted contacts with OPCs in cortical layer V.<sup>132</sup> Synaptic input from local GABAergic interneurons has been shown to dynamically regulate OPC differentiation to oligodendrocytes.<sup>133</sup> OPCs receive strong GABAergic synaptic input from PV, and to a lesser extent from non-PV, interneurons.<sup>132</sup> Notably, the peak neurodevelopmental period of interneuron-OPC connectivity (P10-P14) would thus position interneuron myelination precisely in the window following the initial onset of GABAergic burst firing, but before maturation of high-frequency gamma oscillations.<sup>134</sup> This also closely aligns with the timing of human frontal cortex oligodendrocyte development which plateaus in early adulthood and is highly distinct from white matter development in which oligodendrocytes have already reached their maximum number by ~ 5 years of age.<sup>135</sup> Moreover, in further contrast to white matter, frontal cortex grav matter exhibits a substantial turnover of oligodendrocytes and myelin that persists throughout adulthood.<sup>135</sup> Analogously, rodent studies have demonstrated that OPCs exhibit important distinctions in their physiology, proliferation and differentiation between gray and white matter in rodents.<sup>136</sup> Therefore, regional differences in human OPCs are also not unlikely.

Interestingly, direct contacts of interneurons onto OPCs<sup>137</sup> are only locally distributed, reaching a typical maximum distance of 50–70 µm,<sup>132</sup> which is highly similar to the estimate for the maximal length of OPC processes. An interesting question remains why interneurons have such a restricted spatial localization of their connectivity onto OPCs, since PV cells establish synaptic contacts with pyramidal cells across a distance approximately six times larger.<sup>138</sup> One possibility is that OPCs utilize reciprocal synaptic input to regulate their proliferative drive. Alternatively, it may be that myelination preferentially occurs on proximal axonal segments, in close apposition to the observed localization of OPCs and allowing for rapid differentiation to oligodendrocytes with enhanced myelination plasticity.

# POTENTIAL FUNCTIONS OF INTERNEURON MYELINATION

PV interneurons function to synchronize pyramidal cell ensembles, and thereby generate high-frequency oscillations.<sup>139</sup> Since cortical PV axonal arborization is widely ramified and distributed over

Molecular Psychiatry (2017), 4-12

Table 3. Studies reporting myelination of parvalbumin-expressing int	elination	of parvalbumin-expressing int	erneurons	
Study	Species	Species Brain region	Technique	Conclusion
Micheva et al. <sup>125</sup> Mouse SomMcGee et al. <sup>97</sup> Mouse VisuSomogyi et al. <sup>99</sup> Cat VisuSomogyi et al. <sup>101</sup> Human FronSeress et al. <sup>103</sup> Human FronChung et al. <sup>103</sup> Human FronSeress et al. <sup>103</sup> Human FronHinova-Palova et al. <sup>112</sup> RatPieters et al. <sup>112</sup> RatWouterlood et al. <sup>119</sup> RatBrauer et al. <sup>124</sup> RatKita et al. <sup>123</sup> RatKatsumaru et al. <sup>123</sup> RatHipKatsumaru et al. <sup>122</sup> Ratsumaru et al. <sup>122</sup> RatHipKatsumaru et al. <sup>123</sup> Abbreviations: GABA, gamma-aminobutyric acid;of structurally intact, optically transparent tissue.	Mouse Mouse Cat Human Human Human Rat Rat Rat Rat Rat Rat Rat Rat Rat ansparent ti ansparent ti	Mouse Somatosensory cortex Mouse Visual cortex Cat Visual cortex Cat Visual cortex Human Frontal cortex Human Alppocampus Human Claustrum Rat Visual cortex Rat Visual cortex Rat Hippocampus Rat Hippocampus ( <i>in vitro</i> ) Rat Hippocampus Rat Hippocampus ( <i>in vitro</i> ) Rat Hippocampus ( <i>in vitro</i> )	Array tomography; electron microscopy; immunofluorescence Immunofluorescence Electron microscopy Electron microscopy Electron microscopy; Electron microscopy; Electron microscopy Electron microscopy Electron microscopy Immunofluorescence; electron microscopy Immunofluorescence; electron microscopy Hippocampal cultures; immunofluorescence Electron microscopy Immunofluorescence Chung <i>et al.</i> <sup>100</sup> ) is a recently developed technique which	Micheva <i>et al.</i> <sup>125</sup> Mouse Somatosensory cortex Array tomography: electron microscopy; electron microscopy; electron microscopy in all are PV+ Mich nearly all are PV+ Terestree of several myelinated basket cells Filectron microscopy Filectron microscopy Hinnon Palova <i>et al.</i> <sup>10</sup> Human Hippocampus Filectron microscopy Mouterlood <i>et al.</i> <sup>110</sup> Human Hippocampus Filectron microscopy Mouterlood <i>et al.</i> <sup>113</sup> Mouterlood <i>et al.</i> <sup>113</sup> Mouterlood <i>et al.</i> <sup>113</sup> Mouterlood <i>et al.</i> <sup>113</sup> Mat Hippocampus Filectron microscopy Majority of septohippocampal PV fibers show myelinated PV axons Presence of several myelinated PV neurons Majority of septohippocampal dentate gyrus Immunofluorescence Moterion of PV neurons <i>in vitro</i> Majority of septohippocampal dentate gyrus Immunofluorescence Moterion of PV neurons <i>in vitro</i> Majority presence of myelinated PV neurons Moterion of PV neurons <i>in vitro</i> Majority optically transpartisue.

distances of up to 300 µm,<sup>138</sup> there might be considerable benefits of myelination for optimizing the fidelity of fast action potential transmission. Indeed, computational modeling has suggested a unique contribution of (interneuron) conductance delays in the dynamics of gamma frequency oscillations.<sup>140</sup> Evidence exists that nodes of Ranvier begin forming before the onset of myelination,<sup>117</sup> a mechanism specific for GABAergic neurons, which enhances axonal conduction of action potentials without myelin. Thus, in addition to simply increasing the speed of action potential propagation, myelin could function to ensure the integrity of precisely timed action potentials, as has been proposed for myelinated excitatory axons.<sup>141</sup> Myelin plasticity would then have the potential to support the local synchronization of action potentials necessary for generating high-frequency oscillations.<sup>142</sup> Indeed, myelinated axons exhibit both higher conduction velocities and enhanced long-range coherence.<sup>143</sup> Although non-PV cortical interneuron subtypes (e.g., somatostatin, VIP) exhibit synaptic connectivity across similar distances, <sup>138</sup> their lack of influence in maintaining high-frequency oscillations is consistent with their absence of myelination. Furthermore, the activity-dependence of myelination<sup>144</sup> might permit dynamically regulated influences on the fidelity of fast action potential transmission and high-frequency oscillations.

Furthermore, myelin could provide metabolic and trophic support for energetically costly PV cells. PV cell characteristics, including high-frequency spiking and rapid action potential kinetics, require a particularly high energy utilization through predominantly mitochondrial oxidative phosphorylation.<sup>145</sup> Gamma band synchrony, closely linked to cognition, is highly sensitive to metabolic disruption. Furthermore, compared with pyramidal cells, PV cells exhibit high densities of mitochondria and expression of cytochrome *c* and cytochrome *c* oxidase, proteins crucial for the electron transport chain. Moreover, PV cell-specific disruption of cytochrome oxidase assembly leads to changes in PV cell intrinsic excitability, afferent synaptic input, and gamma/theta oscillations, as well as schizophrenia-related behavioral impairments in sensory gating and social behavior.<sup>146</sup>

During gamma oscillations, peak oxygen consumption approaches the demand observed during seizures and mitochondrial oxidative capacity operates near its functional limit.<sup>145</sup> Metabolic and trophic support conferred by myelination<sup>147,148</sup> might therefore allow PV axons to optimize their energy utilization. Consistent with the importance of myelination in regulating axonal energy metabolism is the considerable discrepancy of mitochondria content (30-fold) in myelinated versus unmyelinated axons.<sup>149</sup> Myelin has been proposed to regulate axonal energy metabolism via the monocarboxylate transporter 1 channel.<sup>147,148</sup> Furthermore, the high-peak oxygen consumption of PV cells during gamma band synchrony could require the additional lactate provided by oligodendrocytes.

Taken together, the electrophysiological dynamics of fastspiking PV interneurons, their dense branching onto pyramidal neurons requiring finely tuned temporally synchronized inhibition, and their high-energy consumption are likely interdependent mechanisms governed by PV interneuron myelination.

# IMPLICATIONS FOR SCHIZOPHRENIA

Both interneuron dysfunction and myelination abnormalities have been independently proposed as important contributors to the underlying pathophysiology of schizophrenia. These mechanisms have each amassed convincing support from postmortem histopathology, *in vivo* imaging and electrophysiology, genetics and neurodevelopment (Table 1). However, neither hypothesis is capable of accounting for the full set of clinical research findings in schizophrenia. In contrast, interneuron myelination brings together both of these models, explains a more comprehensive portion of the existing data, and offers a well-defined falsifiable model.

Impairments of PV interneuron myelination could directly contribute to schizophrenia through several mechanisms. Impaired action potential fidelity, energy restrictions during highly-demanding cognitive tasks, aberrant axonal branching and a higher occurrence of ectopic action potentials could each independently, or in combination, disrupt inhibitory network function. Such changes to PV interneurons would likely result in abnormalities of local gamma synchronization, with a potential further impact on the integrity of long-range thalamocortical and cortico-striatal circuits, and striatal dopamine signaling, ultimately giving rise to schizophrenia symptoms.

In this Perspective, we have proposed the novel hypothesis that altered myelination of PV interneurons might function prominently in the pathophysiology of schizophrenia. However, many questions remain to be answered. At what point during development does interneuron myelination occur and to what extent does this coincide with the clinical symptoms of schizophrenia? Does interneuron myelination vary across brain regions? Is cortical interneuron myelination truly reserved for fast-spiking PV interneurons, or are non-fast-spiking interneurons (e.g., somatostatin, VIP) myelinated as well? How does the plasticity of PV interneuron myelination compare with that of glutamatergic axons? And perhaps most importantly, to what extent might PV interneuron myelination represent an etiological pathophysiology and therapeutic target for schizophrenia?

Future studies to examine the parvalbumin interneuron myelination hypothesis could be approached through a variety of methods. In particular, the most important experiments would include: (a) detailed histological assessments of subtype-specific interneuron axonal myelination in postmortem brain tissue from patients with schizophrenia, (b) corresponding functional studies in rodent models of schizophrenia to directly assess the causality of alterations in parvalbumin interneuron myelination on behavioral and electrophysiological phenotypes, (c) electrophysiological studies of rodent models with temporally-controlled and cell-type specific disruption of myelination and (d) functional genomic studies on the effect of schizophrenia risk variants on (interneuron) myelination, for example, by utilizing human induced pluripotent stem cells or genetically modified mice.

# **CONFLICT OF INTEREST**

The authors declare no conflict of interest.

# ACKNOWLEDGMENTS

Funding was provided by ZonMw Vidi 017.106.384 from the Netherlands Organization for Scientific Research and NeuroBasic PharmaPhenomics consortium to SAK, and the Dutch Technology Foundation STW applied science division of NWO and the Technology Programme of the Ministry of Economic Affairs (Project 12197) to SAK and JS.

#### REFERENCES

- 1 Owen MJ, Sawa A, Mortensen PB. Schizophrenia. Lancet 2016; 388: 86-97.
- 2 Insel TR. Rethinking schizophrenia. Nature 2010; 468: 187-193.
- 3 Carlsson A. The current status of the dopamine hypothesis of schizophrenia. *Neuropsychopharmacology* 1988; 1: 179–186.
- 4 Moghaddam B, Javitt D. From revolution to evolution: the glutamate hypothesis of schizophrenia and its implication for treatment. *Neuropsychopharmacology* 2012; **37**: 4–15.
- 5 Gonzalez-Burgos G, Cho RY, Lewis DA. Alterations in cortical network oscillations and parvalbumin neurons in schizophrenia. *Biol Psychiatry* 2015; 77: 1031–1040.
- 6 Fitzsimmons J, Kubicki M, Shenton ME. Review of functional and anatomical brain connectivity findings in schizophrenia. *Curr Opin Psychiatry* 2013; 26: 172–187.
- 7 Lewis DA, Curley AA, Glausier JR, Volk DW. Cortical parvalbumin interneurons and cognitive dysfunction in schizophrenia. *Trends Neurosci* 2012; 35: 57–67.

- 8 Gonzalez-Burgos G, Hashimoto T, Lewis DA. Alterations of cortical GABA neurons and network oscillations in schizophrenia. *Curr Psychiatry Rep* 2010; 12: 335–344.
- 9 Hashimoto T, Arion D, Unger T, Maldonado-Avilés JG, Morris HM, Volk DW et al. Alterations in GABA-related transcriptome in the dorsolateral prefrontal cortex of subjects with schizophrenia. *Mol Psychiatry* 2008; **13**: 147–161.
- 10 Thompson M, Weickert CS, Wyatt E, Webster MJ. Decreased glutamic acid decarboxylase(67) mRNA expression in multiple brain areas of patients with schizophrenia and mood disorders. J Psychiatr Res 2009; 43: 970–977.
- 11 Woo TU, Walsh JP, Benes FM. Density of glutamic acid decarboxylase 67 messenger RNA-containing neurons that express the N-methyl-D-aspartate receptor subunit NR2A in the anterior cingulate cortex in schizophrenia and bipolar disorder. Arch Gen Psychiatry 2004; 61: 649–657.
- 12 Impagnatiello F. A decrease of reelin expression as a putative vulnerability factor. *Proc Natl Acad Sci USA* 1998; **95**: 15718–15723.
- 13 Guidotti A, Auta J, Davis JM, Di-Giorgi-Gerevini V, Dwivedi Y, Grayson DR *et al.* Decrease in reelin and glutamic acid decarboxylase67 (GAD67) expression in schizophrenia and bipolar disorder: a postmortem brain study. *Arch Gen Psychiatry* 2000; **57**: 1061–1069.
- 14 Curley AA, Arion D, Volk DW, Asafu-Adjei JK, Sampson AR, Fish KN et al. Cortical deficits of glutamic acid decarboxylase 67 expression in schizophrenia: clinical, protein, and cell type-specific features. Am J Psychiatry 2011; 168: 921–929.
- 15 Akbarian S, Kim JJ, Potkin SG, Hagman JO, Tafazzoli A, Bunney WE et al. Gene expression for glutamic acid decarboxylase is reduced without loss of neurons in prefrontal cortex of schizophrenics. Arch Gen Psychiatry 1995; 52: 258–266.
- 16 Volk DW, Austin MC, Pierri JN, Sampson AR, Lewis DA. Decreased glutamic acid decarboxylase67 messenger RNA expression in a subset of prefrontal cortical gamma-aminobutyric acid neurons in subjects with schizophrenia. Arch Gen Psychiatry 2000; 57: 237–245.
- 17 Hashimoto T, Volk DW, Eggan SM, Mirnics K, Pierri JN, Sun Z et al. Gene expression deficits in a subclass of GABA neurons in the prefrontal cortex of subjects with schizophrenia. J Neurosci 2003; 23: 6315–6326.
- 18 Hashimoto T, Bazmi HH, Mirnics K, Wu Q, Sampson AR, Lewis DA. Conserved regional patterns of GABA-related transcript expression in the neocortex of subjects with schizophrenia. Am J Psychiatry 2008; 165: 479–489.
- 19 Mellios N, Huang H-S, Baker SP, Galdzicka M, Ginns E, Akbarian S. Molecular determinants of dysregulated GABAergic gene expression in the prefrontal cortex of subjects with schizophrenia. *Biol Psychiatry* 2009; 65: 1006–1014.
- 20 Fung SJ, Webster MJ, Sivagnanasundaram S, Duncan C, Elashoff M, Weickert CS. Expression of interneuron markers in the dorsolateral prefrontal cortex of the developing human and in schizophrenia. Am J Psychiatry 2010; 167: 1479–1488.
- 21 Glausier JR, Fish KN, Lewis DA. Altered parvalbumin basket cell inputs in the dorsolateral prefrontal cortex of schizophrenia subjects. *Mol Psychiatry* 2014; **19**: 30–36.
- 22 Woo TU, Miller JL, Lewis DA. Schizophrenia and the parvalbumin-containing class of cortical local circuit neurons. *Am J Psychiatry* 1997; **154**: 1013–1015.
- 23 Beasley CL, Zhang ZJ, Patten I, Reynolds GP. Selective deficits in prefrontal cortical GABAergic neurons in schizophrenia defined by the presence of calcium-binding proteins. *Biol Psychiatry* 2002; **52**: 708–715.
- 24 Tooney PA, Chahl LA. Neurons expressing calcium-binding proteins in the prefrontal cortex in schizophrenia. *Prog Neuropsychopharmacol Biol Psychiatry* 2004; 28: 273–278.
- 25 Pantazopoulos H, Woo TU, Lim MP, Lange N, Berretta S. Extracellular matrix-glial abnormalities in the amygdala and entorhinal cortex of subjects diagnosed with schizophrenia. Arch Gen Psychiatry 2010; 67: 155–166.
- 26 Donato F, Rompani SB, Caroni P. Parvalbumin-expressing basket-cell network plasticity induced by experience regulates adult learning. *Nature* 2013; 504: 272–276.
- 27 Brown JA, Ramikie TS, Schmidt MJ, Báldi R, Garbett K, Everheart MG et al. Inhibition of parvalbumin-expressing interneurons results in complex behavioral changes. *Mol Psychiatry* 2015; 20: 1499–1507.
- 28 Frankle WG, Cho RY, Prasad KM, Mason NS, Paris J, Himes ML et al. In vivo measurement of GABA transmission in healthy subjects and schizophrenia patients. Am J Psychiatry 2015; 172: 1148–1159.
- 29 Cardin JA, Carlén M, Meletis K, Knoblich U, Zhang F, Deisseroth K et al. Driving fast-spiking cells induces gamma rhythm and controls sensory responses. *Nature* 2009; 459: 663–667.
- 30 Sohal VS, Zhang F, Yizhar O, Deisseroth K. Parvalbumin neurons and gamma rhythms enhance cortical circuit performance. *Nature* 2009; **459**: 698–702.
- 31 Hormuzdi SG, Pais I, LeBeau FE, Towers SK, Rozov A, Buhl EH et al. Impaired electrical signaling disrupts gamma frequency oscillations in connexin 36-deficient mice. *Neuron* 2001; **31**: 487–495.
- 32 Traub RD, Kopell N, Bibbig A, Buhl EH, LeBeau FE, Whittington MA. Gap junctions between interneuron dendrites can enhance synchrony of gamma oscillations in distributed networks. *J Neurosci* 2001; **21**: 9478–9486.

- 33 Whittington MA, Traub RD, Jefferys JG. Synchronized oscillations in interneuron networks driven by metabotropic glutamate receptor activation. *Nature* 1995; 373: 612–615.
- 34 Senkowski D, Gallinat J. Dysfunctional prefrontal gamma-band oscillations reflect working memory and other cognitive deficits in schizophrenia. *Biol Psychiatry* 2015; **77**: 1010–1019.
- 35 Uhlhaas PJ, Singer W. Oscillations and neuronal dynamics in schizophrenia: the search for basic symptoms and translational opportunities. *Biol Psychiatry* 2014; 77: 1001–1009.
- 36 Pocklington AJ, Rees E, Walters JTR, Han J, Kavanagh DH, Chambert KD et al. Novel findings from CNVs implicate inhibitory and excitatory signaling complexes in schizophrenia. *Neuron* 2015; 86: 1203–1214.
- 37 Karayiorgou M, Morris MA, Morrow B, Shprintzen RJ, Goldberg R, Borrow J et al. Schizophrenia susceptibility associated with interstitial deletions of chromosome 22q11. Proc Natl Acad Sci USA 1995; 92: 7612–7616.
- 38 Lindsay EA, Morris MA, Gos A, Nestadt G, Wolyniec PS, Lasseter VK et al. Schizophrenia and chromosomal deletions within 22q11.2. Am J Hum Genet 1995; 56: 1502–1503.
- 39 Ellegood J, Markx S, Lerch JP, Steadman PE, Genç C, Provenzano F *et al.* Neuroanatomical phenotypes in a mouse model of the 22q11.2 microdeletion. *Mol Psychiatry* 2014; **19**: 99–107.
- 40 Chun S, Westmoreland JJ, Bayazitov IT, Eddins D, Pani AK, Smeyne RJ et al. Specific disruption of thalamic inputs to the auditory cortex in schizophrenia models. Science 2014; 344: 1178–1182.
- 41 Sigurdsson T, Stark KL, Karayiorgou M, Gogos JA, Gordon JA. Impaired hippocampal–prefrontal synchrony in a genetic mouse model of schizophrenia. *Nature* 2010; **464**: 763–767.
- 42 Meechan DW, Tucker ES, Maynard TM, LaMantia A-S. Diminished dosage of 22q11 genes disrupts neurogenesis and cortical development in a mouse model of 22q11 deletion/DiGeorge syndrome. *Proc Natl Acad Sci USA* 2009; **106**: 16434–16445.
- 43 Rosen AM, Spellman T, Gordon JA. Electrophysiological endophenotypes in rodent models of schizophrenia and psychosis. *Biol Psychiatry* 2015; 77: 1041–1049.
- 44 White T, Magnotta VA, Bockholt HJ, Williams S, Wallace S, Ehrlich S *et al.* Global white matter abnormalities in schizophrenia: a multisite diffusion tensor imaging study. *Schizophr Bull* 2011; **37**: 222–232.
- 45 Alvarado-Alanis P, León-Ortiz P, Reyes-Madrigal F, Favila R, Rodríguez-Mayoral O, Nicolini H et al. Abnormal white matter integrity in antipsychotic-naïve first-episode psychosis patients assessed by a DTI principal component analysis. *Schizophr Res* 2015; **162**: 14–21.
- 46 Cheung V, Cheung C, McAlonan GM, Deng Y, Wong JG, Yip L et al. A diffusion tensor imaging study of structural dysconnectivity in never-medicated, firstepisode schizophrenia. Psychol Med 2008; 38: 877–885.
- 47 Filippi M, Canu E, Gasparotti R, Agosta F, Valsecchi P, Lodoli G et al. Patterns of brain structural changes in first-contact, antipsychotic drug-naive patients with schizophrenia. Am J Neuroradiol 2013; 35: 30–37.
- 48 Gasparotti R, Valsecchi P, Carletti F, Galluzzo A, Liserre R, Cesana B et al. Reduced fractional anisotropy of corpus callosum in first-contact, antipsychotic drug-naive patients with schizophrenia. Schizophr Res 2009; 108: 41–48.
- 49 Guo W, Liu F, Liu Z, Gao K, Xiao C, Chen H *et al.* Right lateralized white matter abnormalities in first-episode, drug-naive paranoid schizophrenia. *Neurosci Lett* 2012; **531**: 5–9.
- 50 Pérez-Iglesias R, Tordesillas-Gutiérrez D, Barker GJ, McGuire PK, Roiz-Santiañez R, Mata I et al. White matter defects in first episode psychosis patients: a voxelwise analysis of diffusion tensor imaging. *Neuroimage* 2010; **49**: 199–204.
- 51 Mandl RCW, Rais M, van Baal GCM, van Haren NEM, Cahn W, Kahn RS et al. Altered white matter connectivity in never-medicated patients with schizophrenia. Hum Brain Mapp 2013; 34: 2353–2365.
- 52 Melicher T, Horacek J, Hlinka J, Spaniel F, Tintera J, Ibrahim I *et al.* White matter changes in first episode psychosis and their relation to the size of sample studied: a DTI study *et al.* White matter abnormalities in schizophrenia and schizotypal personality disorder. *Schizophr Bull* 2015; **41**: 300–310.
- 53 Lener MS, Wong E, Tang CY, Byne W, Goldstein KE, Blair NJ et al. White matter abnormalities in schizophrenia and schizotypal personality disorder. Schizophr Bull 2015; 41: 300–310.
- 54 Lyu H, Hu M, Eyler LT, Jin H, Wang J, Ou J et al. Regional white matter abnormalities in drug-naive, first-episode schizophrenia patients and their healthy unaffected siblings. Aust N Z J Psychiatry 2015; **49**: 246–254.
- 55 Liu X, Lai Y, Wang X, Hao C, Chen L, Zhou Z et al. A combined DTI and structural MRI study in medicated-naïve chronic schizophrenia. *Magn Reson Imaging* 2014; 32: 1–8.
- 56 Sun H, Lui S, Yao L, Deng W, Xiao Y, Zhang W et al. Two patterns of white matter abnormalities in medication-naive patients with first-episode schizophrenia revealed by diffusion tensor imaging and cluster analysis. JAMA Psychiatry 2015; 72: 678–686.

- 57 Yao L, Lui S, Deng W, Wu M, Chen L, Xiao Y *et al.* Association of white matter deficits with clinical symptoms in antipsychotic-naive first-episode schizo-phrenia: an optimized VBM study using 3 T. *MAGMA* 2014; **27**: 283–290.
- 58 Wang Q, Cheung C, Deng W, Li M, Huang C, Ma X et al. White-matter microstructure in previously drug-naive patients with schizophrenia after 6 weeks of treatment. Psychol Med 2013; 43: 2301–2309.
- 59 Do KQ, Cuenod M, Hensch TK. Targeting oxidative stress and aberrant critical period plasticity in the developmental trajectory to schizophrenia. *Schizophr Bull* 2015; **41**: 835–846.
- 60 Hermoye L, Saint-Martin C, Cosnard G, Lee S-K, Kim J, Nassogne M-C et al. Pediatric diffusion tensor imaging: normal database and observation of the white matter maturation in early childhood. *Neuroimage* 2006; 29: 493–504.
- 61 Lebel C, Walker L, Leemans A, Phillips L, Beaulieu C. Microstructural maturation of the human brain from childhood to adulthood. *Neuroimage* 2008; 40: 1044–1055.
- 62 Lebel C, Gee M, Camicioli R, Wieler M, Martin W, Beaulieu C. Diffusion tensor imaging of white matter tract evolution over the lifespan. *Neuroimage* 2012; 60: 340–352.
- 63 Miller DJ, Duka T, Stimpson CD, Schapiro SJ, Baze WB, McArthur MJ et al. Prolonged myelination in human neocortical evolution. *Proc Natl Acad Sci USA* 2012; **109**: 16480–16485.
- 64 Tkachev D, Mimmack ML, Ryan MM, Wayland M, Freeman T, Jones PB et al. Oligodendrocyte dysfunction in schizophrenia and bipolar disorder. *Lancet* 2003; 362: 798–805.
- 65 Hakak Y, Walker JR, Li C, Wong WH, Davis KL, Buxbaum JD et al. Genome-wide expression analysis reveals dysregulation of myelination-related genes in chronic schizophrenia. Proc Natl Acad Sci USA 2001; 98: 4746–4751.
- 66 Barley K, Dracheva S, Byne W. Subcortical oligodendrocyte- and astrocyteassociated gene expression in subjects with schizophrenia, major depression and bipolar disorder. *Schizophr Res* 2009; **112**: 54–64.
- 67 Katsel P, Davis KL, Gorman JM, Haroutunian V. Variations in differential gene expression patterns across multiple brain regions in schizophrenia. *Schizophr Res* 2005; **77**: 241–252.
- 68 Aberg K, Saetre P, Jareborg N, Jazin E. Human QKI, a potential regulator of mRNA expression of human oligodendrocyte-related genes involved in schizophrenia. *Proc Natl Acad Sci USA* 2006; **103**: 7482–7487.
- 69 Schmitt A, Wilczek K, Blennow K, Maras A, Jatzko A, Petroianu G et al. Altered thalamic membrane phospholipids in schizophrenia: a postmortem study. *Biol Psychiatry* 2004; 56: 41–45.
- 70 Matthews PR, Eastwood SL, Harrison PJ. Reduced myelin basic protein and actinrelated gene expression in visual cortex in schizophrenia. *PLoS One* 2012; 7: e38211.
- 71 Aston C, Jiang L, Sokolov BP. Microarray analysis of postmortem temporal cortex from patients with schizophrenia. J Neurosci Res 2004; 77: 858–866.
- 72 Harris LW, Lockstone HE, Khaitovich P, Weickert CS, Webster MJ, Bahn S. Gene expression in the prefrontal cortex during adolescence: implications for the onset of schizophrenia. *BMC Med Genomics* 2009; 2: 28.
- 73 Hof PR, Haroutunian V, Copland C, Davis KL, Buxbaum JD. Molecular and cellular evidence for an oligodendrocyte abnormality in schizophrenia. *Neurochem Res* 2002; 27: 1193–1200.
- 74 Hof PR, Haroutunian V, Friedrich VL, Byne W, Buitron C, Perl DP et al. Loss and altered spatial distribution of oligodendrocytes in the superior frontal gyrus in schizophrenia. Biol Psychiatry 2003; 53: 1075–1085.
- 75 Uranova NA, Kolomeets NS, Vigreva OV, Zimina IS, Rachmanova VI, Orlovskaya DD. Ultrastructural pathology of myelinated fibers in schizophrenia. *Zh Nevrol Psikhiatr Im S S Korsakova* 2013; **113**: 63–69.
- 76 Uranova NA, Vostrikov VM, Vikhreva OV, Zimina IS, Kolomeets NS, Orlovskaya DD. The role of oligodendrocyte pathology in schizophrenia. Int J Neuropsychopharmacol 2007; 10: 537–545.
- 77 Vostrikov VM, Uranova NA, Rakhmanova VI, Orlovskaia DD. Lowered oligodendroglial cell density in the prefrontal cortex in schizophrenia. *Zh Nevrol Psikhiatr Im S S Korsakova* 2004; **104**: 47–51.
- 78 Uranova NA, Vostrikov VM, Orlovskaya DD, Rachmanova VI. Oligodendroglial density in the prefrontal cortex in schizophrenia and mood disorders: a study from the Stanley Neuropathology Consortium. *Schizophr Res* 2004; 67: 269–275.
- 79 Schmitt A, Steyskal C, Bernstein H-G, Schneider-Axmann T, Parlapani E, Schaeffer EL et al. Stereologic investigation of the posterior part of the hippocampus in schizophrenia. Acta Neuropathol 2009; **117**: 395–407.
- 80 Kerns D, Vong GS, Barley K, Dracheva S, Katsel P, Casaccia P *et al.* Gene expression abnormalities and oligodendrocyte deficits in the internal capsule in schizophrenia. *Schizophr Res* 2010; **120**: 150–158.
- 81 Williams MR, Marsh R, Macdonald CD, Jain J, Pearce RKB, Hirsch SR et al. Neuropathological changes in the nucleus basalis in schizophrenia. Eur Arch Psychiatry Clin Neurosci 2013; 263: 485–495.

- 82 Byne W, Tatusov A, Yiannoulos G, Vong GS, Marcus S. Effects of mental illness and aging in two thalamic nuclei. *Schizophr Res* 2008; **106**: 172–181.
- 83 Mauney SA, Pietersen CY, Sonntag KC, Woo TU. Differentiation of oligodendrocyte precursors is impaired in the prefrontal cortex in schizophrenia. *Schizophr Res* 2015; **169**: 374–380.
- 84 Williams MR, Galvin K, O'Sullivan B, MacDonald CD, Ching EWK, Turkheimer F et al. Neuropathological changes in the substantia nigra in schizophrenia but not depression. Eur Arch Psychiatry Clin Neurosci 2014; 264: 285–296.
- 85 Williams MR, Hampton T, Pearce RKB, Hirsch SR, Ansorge O, Thom M *et al.* Astrocyte decrease in the subgenual cingulate and callosal genu in schizophrenia. *Eur Arch Psychiatry Clin Neurosci* 2013; **263**: 41–52.
- 86 Chambers JS, Perrone-Bizzozero NI. Altered myelination of the hippocampal formation in subjects with schizophrenia and bipolar disorder. *Neurochem Res* 2004; 29: 2293–2302.
- 87 Bernstein H-G, Smalla K-H, Dürrschmidt D, Keilhoff G, Dobrowolny H, Steiner J et al. Increased density of prohibitin-immunoreactive oligodendrocytes in the dorsolateral prefrontal white matter of subjects with schizophrenia suggests extraneuronal roles for the protein in the disease. *Neuromolecular Med* 2012; 14: 270–280.
- 88 Williams MR, Harb H, Pearce RKB, Hirsch SR, Maier M. Oligodendrocyte density is changed in the basolateral amygdala in schizophrenia but not depression. *Schizophr Res* 2013; **147**: 402–403.
- 89 Miyakawa T, Sumiyoshi S, Deshimaru M, Suzuki T, Tomonari H. Electron microscopic study on schizophrenia. Mechanism of pathological changes. Acta Neuropathol 1972; 20: 67–77.
- 90 Uranova NA, Vikhreva OV, Rachmanova VI, Orlovskaya DD. Ultrastructural alterations of myelinated fibers and oligodendrocytes in the prefrontal cortex in schizophrenia: a postmortem morphometric study. *Schizophr Res Treatment* 2011; **2011**: 325789.
- 91 Toritsuka M, Makinodan M, Kishimoto T. Social experience-dependent myelination: an implication for psychiatric disorders. *Neural Plast* 2015; **2015**: 465345.
- 92 Duncan LE, Holmans PA, Lee PH, O'Dushlaine CT, Kirby AW, Smoller JW et al. Pathway analyses implicate glial cells in schizophrenia. PLoS One 2014; 9: e89441.
- 93 Goudriaan A, de Leeuw C, Ripke S, Hultman CM, Sklar P, Sullivan PF et al. Specific glial functions contribute to schizophrenia susceptibility. *Schizophr Bull* 2014; 40: 925–935.
- 94 Xu K, Schadt EE, Pollard KS, Roussos P, Dudley JT. Genomic and network patterns of schizophrenia genetic variation in human evolutionary accelerated regions. *Mol Biol Evol* 2015; **32**: 1148–1160.
- 95 Chavarria-Siles I, White T, de Leeuw C, Goudriaan A, Lips E, Ehrlich S et al. Myelination-related genes are associated with decreased white matter integrity in schizophrenia. *Eur J Hum Genet* 2016; 24: 381–386.
- 96 Ripke S, Neale BM, Corvin A, Walters JTR, Farh K-H, Holmans PA *et al.* Biological insights from 108 schizophrenia-associated genetic loci. *Nature* 2014; **511**: 421–427.
- 97 McGee AW, Yang Y, Fischer QS, Daw NW, Strittmatter SM. Experience-driven plasticity of visual cortex limited by myelin and Nogo receptor. *Science* 2005; 309: 2222–2226.
- 98 Somogyi P, Soltész I. Immunogold demonstration of GABA in synaptic terminals of intracellularly recorded, horseradish peroxidase-filled basket cells and clutch cells in the cat's visual cortex. *Neuroscience* 1986; 19: 1051–1065.
- 99 Somogyi P, Kisvárday ZF, Martin KA, Whitteridge D. Synaptic connections of morphologically identified and physiologically characterized large basket cells in the striate cortex of cat. *Neuroscience* 1983; **10**: 261–294.
- 100 Somogyi P, Freund TF, Wu JY, Smith AD. The section-Golgi impregnation procedure. 2. Immunocytochemical demonstration of glutamate decarboxylase in Golgi-impregnated neurons and in their afferent synaptic boutons in the visual cortex of the cat. *Neuroscience* 1983; **9**: 475–490.
- 101 Chung K, Wallace J, Kim S-Y, Kalyanasundaram S, Andalman AS, Davidson TJ et al. Structural and molecular interrogation of intact biological systems. *Nature* 2013; **497**: 332–337.
- 102 Ong WY, Yeo TT, Balcar VJ, Garey LJ. A light and electron microscopic study of GAT-1-positive cells in the cerebral cortex of man and monkey. J Neurocytol 1998; 27: 719–730.
- 103 Seress L, Gulyás AI, Ferrer I, Tunon T, Soriano E, Freund TF. Distribution, morphological features, and synaptic connections of parvalbumin- and calbindin D28k-immunoreactive neurons in the human hippocampal formation. J Comp Neurol 1993; 337: 208–230.
- 104 Hinova-Palova DV, Edelstein L, Landzhov BV, Braak E, Malinova LG, Minkov M *et al.* Parvalbumin-immunoreactive neurons in the human claustrum. *Brain Struct Funct* 2014; **219**: 1813–1830.
- 105 Sloper JJ, Powell TP. A study of the axon initial segment and proximal axon of neurons in the primate motor and somatic sensory cortices. *Philos Trans R Soc Lond B Biol Sci* 1979; **285**: 173–197.

- 106 Hendry SH, Houser CR, Jones EG, Vaughn JE. Synaptic organization of immunocytochemically identified GABA neurons in the monkey sensory-motor cortex. *J Neurocytol* 1983; **12**: 639–660.
- 107 DeFelipe J, Jones EG. Vertical organization of gamma-aminobutyric acid-accumulating intrinsic neuronal systems in monkey cerebral cortex. J Neurosci 1985; 5: 3246–3260.
- 108 DeFelipe J, Hendry SH, Jones EG. A correlative electron microscopic study of basket cells and large GABAergic neurons in the monkey sensory-motor cortex. *Neuroscience* 1986; **17**: 991–1009.
- 109 Takasu N, Nakatani T, Arikuni T, Kimura H. Immunocytochemical localization of gammaaminobutyric acid in the hypoglossal nucleus of the macaque monkey, Macaca fuscata: a light and electron microscopic study. J Comp Neurol 1987; 263: 42–53.
- 110 Ralston DD, Milroy AM. Inhibitory synaptic input to identified rubrospinal neurons in Macaca fascicularis: an electron microscopic study using a combined immuno-GABA-gold technique and the retrograde transport of WGA-HRP. *J Comp Neurol* 1992; **320**: 97–109.
- 111 Jinno S, Klausberger T, Marton LF, Dalezios Y, Roberts JDB, Fuentealba P et al. Neuronal diversity in GABAergic long-range projections from the hippocampus. J Neurosci 2007; 27: 8790–8804.
- 112 Peters A, Proskauer CC. Smooth or sparsely spined cells with myelinated axons in rat visual cortex. *Neuroscience* 1980; **5**: 2079–2092.
- 113 De Biasi S, Amadeo A, Arcelli P, Frassoni C, Spreafico R. Postnatal development of GABA-immunoreactive terminals in the reticular and ventrobasal nuclei of the rat thalamus: a light and electron microscopic study. *Neuroscience* 1997; 76: 503–515.
- 114 Conti F, Zuccarello LV, Barbaresi P, Minelli A, Brecha NC, Melone M. Neuronal, glial, and epithelial localization of gamma-aminobutyric acid transporter 2, a high-affinity gamma-aminobutyric acid plasma membrane transporter, in the cerebral cortex and neighboring structures. J Comp Neurol 1999; 409: 482–494.
- 115 Gartner U, Hartig CAW, Brauer K, Brückner G, Arendt T. Electron microscopic evidence for different myelination of rat septohippocampal fibres. *Neuroreport* 2001; **12**: 17–20.
- 116 Gartner U, Brauer K, Hartig W, Brückner G, Arendt T. Different myelination of rat septohippocampal fibres as revealed by immunofluorescence double-labelling. *Brain Res* 2000; 878: 188–193.
- 117 Freeman SA, Desmazières A, Simonnet J, Gatta M, Pfeiffer F. Acceleration of conduction velocity linked to clustering of nodal components precedes myelination. *Proc Natl Acad Sci USA* 2015; **112**: E321–E328.
- 118 Mize RR. Immunocytochemical localization of gamma-aminobutyric acid (GABA) in the cat superior colliculus. *J Comp Neurol* 1988; **276**: 169–187.
- 119 Wouterlood FG, Härtig W, Brückner G, Witter MP. Parvalbumin-immunoreactive neurons in the entorhinal cortex of the rat: localization, morphology, connectivity and ultrastructure. J Neurocytol 1995; 24: 135–153.
- 120 Sawyer SF, Martone ME, Groves PM. A GABA immunocytochemical study of rat motor thalamus: light and electron microscopic observations. *Neuroscience* 1991; 42: 103–124.
- 121 Roberts RC. An electron microscopic study of GABAergic neurons and terminals in the central nucleus of the inferior colliculus of the rat. *J Neurocytol* 1987; **16**: 333–345.
- 122 Katsumaru H, Kosaka T, Heizmann CW, Hama K. Immunocytochemical study of GABAergic neurons containing the calcium-binding protein parvalbumin in the rat hippocampus. *Exp Brain Res* 1988; **72**: 347–362.
- 123 Kita H, Kosaka T, Heizmann CW. Parvalbumin-immunoreactive neurons in the rat neostriatum: a light and electron microscopic study. *Brain Res* 1990; **536**: 1–15.
- 124 Brauer K, Härtig W, Gärtner U, Brückner G, Arendt T. Different myelination of rat septohippocampal fibres as revealed by immunofluorescence double-labelling. *Brain Res* 2000; 878: 188–193.
- 125 Micheva KD, Wolman D, Mensh BD, Pax E, Buchanan J, Smith SJ et al. A large fraction of neocortical myelin ensheathes axons of local inhibitory neurons. *Elife* 2016; 5: e15784.
- 126 Aoki C, Pickel VM. Neuropeptide Y in cortex and striatum. Ultrastructural distribution and coexistence with classical neurotransmitters and neuropeptides. *Ann N Y Acad Sci* 1990; 611: 186–205.
- 127 Cipolloni PB, Keller A. Thalamocortical synapses with identified neurons in monkey primary auditory cortex: a combined Golgi/EM and GABA/peptide immunocytochemistry study. *Brain Res* 1989; **492**: 347–355.
- 128 Mizukawa K, McGeer PL, Vincent SR, McGeer EG. The distribution of somatostatin-immunoreactive neurons and fibers in the rat cerebral cortex: light and electron microscopic studies. *Brain Res* 1987; **426**: 28–36.
- 129 De Lima AD, Morrison JH. Ultrastructural analysis of somatostatinimmunoreactive neurons and synapses in the temporal and occipital cortex of the macaque monkey. J Comp Neurol 1989; 283: 212–227.
- 130 DiFiglia M, Aronin N. Ultrastructural features of immunoreactive somatostatin neurons in the rat caudate nucleus. *J Neurosci* 1982; **2**: 1267–1274.

- 131 Hinova-Palova DV, Landzhov B, Dzhambazova E, Minkov M, Edelstein L, Malinova L et al. Neuropeptide Y immunoreactivity in the cat claustrum: A light- and electronmicroscopic investigation. J Chem Neuroanat 2014; 61–62: 107–119.
- 132 Orduz D, Maldonado PP, Balia M, Vélez-Fort M, de Sars V, Yanagawa Y et al. Interneurons and oligodendrocyte progenitors form a structured synaptic network in the developing neocortex. *Elife* 2015; 4: e06953.
- 133 Zonouzi M, Scafidi J, Li P, McEllin B, Edwards J, Dupree JL et al. GABAergic regulation of cerebellar NG2 cell development is altered in perinatal white matter injury. Nat Neurosci 2015; 18: 674–682.
- 134 Le Magueresse C, Monyer H. GABAergic interneurons shape the functional maturation of the cortex. *Neuron* 2013; **77**: 388–405.
- 135 Yeung MSY, Zdunek S, Bergmann O, Bernard S, Salehpour M, Alkass K et al. Dynamics of oligodendrocyte generation and myelination in the human brain. *Cell* 2014; **159**: 766–774.
- 136 Hill RA, Patel KD, Medved J, Reiss AM, Nishiyama A. NG2 cells in white matter but not gray matter proliferate in response to PDGF. J Neurosci 2013; 33: 14558–14566.
- 137 Lin S, Bergles DE. Synaptic signaling between GABAergic interneurons and oligodendrocyte precursor cells in the hippocampus. *Nat Neurosci* 2004; 7: 24–32.
- 138 Fino E, Packer AM, Yuste R. The logic of inhibitory connectivity in the neocortex. *Neuroscientist* 2013; **19**: 228–237.
- 139 Hu H, Gan J, Jonas P. Fast-spiking, parvalbumin+ GABAergic interneurons: from cellular design to microcircuit function. *Science* 2014; **345**: 1255263.
- 140 Jadi MP, Margarita Behrens M, Sejnowski TJ. Abnormal gamma oscillations in N-Methyl-D-Aspartate receptor hypofunction models of schizophrenia. *Biol Psychiatry* 2015; **79**: 716–726.
- 141 Yamazaki Y, Fujiwara H, Kaneko K, Hozumi Y, Xu M, Ikenaka K *et al.* Short- and long-term functional plasticity of white matter induced by oligodendrocyte depolarization in the hippocampus. *Glia* 2014; **62**: 1299–1312.
- 142 Pajevic S, Basser PJ, Fields RD. Role of myelin plasticity in oscillations and synchrony of neuronal activity. *Neuroscience* 2014; **276**: 135–147.
- 143 Salami M, Itami C, Tsumoto T, Kimura F. Change of conduction velocity by regional myelination yields constant latency irrespective of distance between thalamus and cortex. *Proc Natl Acad Sci USA* 2003; **100**: 6174–6179.

- 144 Fields RD. A new mechanism of nervous system plasticity: activity-dependent myelination. *Nat Rev Neurosci* 2015; **16**: 756–767.
- 145 Kann O, Papageorgiou IE, Draguhn A. Highly energized inhibitory interneurons are a central element for information processing in cortical networks. J Cereb Blood Flow Metab 2014; 34: 1270–1282.
- 146 Inan M, Zhao M, Manuszak M, Karakaya C, Rajadhyaksha A, Pickel VM et al. Energy deficit in parvalbumin neurons leads to circuit dysfunction, impaired sensory gating and social disability. Neurobiol Dis 2016; 93: 35–46.
- 147 Lee Y, Morrison BM, Li Y, Lengacher S, Farah MH, Hoffman PN et al. Oligodendroglia metabolically support axons and contribute to neurodegeneration. *Nature* 2012; 487: 443–448.
- 148 Fünfschilling U, Supplie LM, Mahad D, Boretius S, Saab AS, Edgar J *et al.* Glycolytic oligodendrocytes maintain myelin and long-term axonal integrity. *Nature* 2012; **485**: 517–521.
- 149 Kasthuri N, Hayworth KJ, Berger DR, Schalek RL, Conchello JA, Knowles-Barley S *et al.* Saturated reconstruction of a volume of neocortex. *Cell* 2015; **162**: 648–661.
- 150 Pietersen CY, Mauney SA, Kim SS, Passeri E, Lim MP, Rooney RJ et al. Molecular profiles of parvalbumin-immunoreactive neurons in the superior temporal cortex in schizophrenia. J Neurogenet 2014; 28: 70–85.
- 151 Hu H, Jonas P. A supercritical density of Na(+) channels ensures fast signaling in GABAergic interneuron axons. *Nat Neurosci* 2014; **17**: 686–693.

This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit http://creativecommons.org/licenses/ by/4.0/

© The Author(s) 2017